## **Experimental Procedures**

## Samples

<u>Single fluorescent color control samples</u> – use 200  $\mu$ l for single color control. You can mix non-Syto single color controls.

Experimental samples – use 500 µl and stain according to following protocol.

## <u>Materials</u>

- 01. anti-CD45 PerCP : BD (Cat. 347464), 5X
- 02. anti-CD14 PE: Caltag
- 03. anti-CD16 PE: Caltag
- 04. Syto-13: Molecular Probes (Cat.)
- 05. BD FACS-lyse
- 06. Phosphate buffered saline without  $Ca^{2+}/Mg^{2+}$  (PBS)
- 07. Lavender Top Vacutainer
- 08. Holder
- 09. Needle

## **Cell preparation**

Draw blood in anticoagulant such as heparanized (green top) or K3 EDTA (lavender top) vacutainer tubes. Assume 5-10 x10<sup>6</sup> cells per ml. Tubes should be processed immediately, as CD45 levels will change at RT. Aliquot appropriate amount to 15 cc conical centrifuge tubes. Place on ice prior to staining.

- 01. Stain CD45 PerCP (1:5) and CD16PE (1:30) or CD14PE (1:20) 10' on ice
- 02. Add 10X volume of 1X BD FACS-lyse at RT for 10' to lyse RBC and fix cells
- 03. Centrifuge 300xg 10'
- 04. Resuspend pellet in PBS at 10<sup>6</sup> cells/ml and filter 70 um mesh
- 05. Centrifuge 300xg rpm 10'
- 06. Resuspend 5-10 x10<sup>7</sup> cells per ml in PBS + 200 nM Syto13 and run on ImageStream